

E.04. Involvement of interleukin-1 receptors in sickness responses to lipopolysaccharide

○Takashi Matsuwaki^{1,2}, Kiseko Shionoya¹, Robert Ihnatko¹, Anna Eskilsson¹, Shigeru Kakuta³, Sylvie Dufour⁴, Markus Schwaninger⁵, Ari Waisman⁶, Werner Müller⁷, Emmanuel Pinteaux⁷, David Engblom¹, Anders Blomqvist¹

¹Department of Clinical and Experimental Medicine, Linköping University, Sweden

²Department of Veterinary Physiology, The University of Tokyo, Japan

³Department of Biomedical Science, The University of Tokyo, Japan

⁴Institut Curie/CNRS UMR144, Paris, France

⁵Institute of Experimental and Clinical Pharmacology and Toxicology, University of Lübeck, Germany

⁶Institute for Molecular Medicine, University Medical Center of the Johannes Gutenberg University Mainz, Germany

⁷Faculty of Biology, Medicine and Health, University of Manchester, United Kingdom

Background & Aim

Peripheral immune stimuli are signaled to the central nervous system to induce various kinds of sickness responses, such as fever, anorexia and stress hormone release. The proinflammatory cytokine interleukin-1 β (IL-1 β) has been suggested to play a major role in this signal transduction. However, there are some conflicting reports about the effect of inhibition/deletion of the transducing receptor for IL-1 β , the IL-1 type I receptor (IL-1R1). In the present study, we first re-examined this issue by evaluating the effect of global gene deletion of IL-1R1 on lipopolysaccharide (LPS) induced anorexia, stress hormone release and fever. We then examined the contribution of tumor necrosis factor- α (TNF α) and IL-6 receptor signaling to the temperature response. Furthermore, by using animals with cell/tissue specific deletion of IL-1R1, we examined the role of IL-1R1s in brain endothelial cells, neural cells or peripheral nerves related to IL-1R1 dependent fever.

Materials & Methods

Animals

◇Global IL-1R1 knockout (KO) : B6.129S7-Il1r1tm1Imx/J

◇Mice with tissue-specific deletions of IL-1R1:

created by crossing mice possessing loxP sites flanking exon 5 of the *Il1r1* gene with mice expressing Cre recombinase under

-*Nestin* promoter (expressed in the central and peripheral nervous system)

-endogenous *Trpv1* locus (expressed by nociceptors in primary sensory ganglia)

-*human tissue plasminogen activator (HtPa)* promoter (expressed by neural crest derivatives)

or mice expressing

-tamoxifen-inducible CreER^{T2} under the *Slco1c1* promoter (expressed in the cerebrovascular endothelium)

Temperature recording

Temperature data were sampled every minute for 9 hours by a transmitter (E-Mitter, Mini Mitter)

Injection of LPS or saline and taking samples

WT and KO mice were injected ip with LPS from *Escherichia coli* (serotype O111:B4; 120 μ g/kg, ip) or saline. One, 3 and 6 hours after injection, brains and blood were taken for analyses.

Immunohistochemistry

Brain sections were cut at 40 μ m on a freezing microtome and incubated in primary antibody solution (anti-c-Fos antibody, Millipore, 1:10000; Millipore) for 60 hours at 4°C, treated with secondary antibody for 2 hours, and then with Avidin-Biotin complex solution for 2 hours at room temperature.

Acknowledgment

Funding: Japan Society for the Promotion of Science (15K18800 to TM), Swedish Medical Research Council (#20725 to DE and #07879 to AB), European Research Council (ERC-starting grant to DE), Knut and Alice Wallenberg foundation (DE), Swedish Brain Foundation (DE and AB), Swedish Cancer Foundation (#213/692 to AB), County Council of Östergötland (DE and AB).

We thank Dr. Yoichiro Iwakura for providing IL-1R1 KO mice.

Results

1. IL-1R1 deletion attenuates the febrile response to LPS

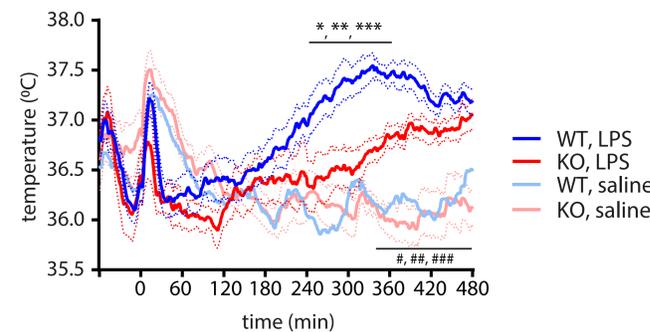


Figure 1. Effect of global IL-1R1 deletion on the febrile response to LPS. LPS (120 μ g/kg) or saline was injected intraperitoneally at 0h. *, **, and *** indicate $P < 0.05$, 0.01, and 0.001 between WT mice and IL-1R1 KO mice treated with LPS, and #, ##, and ### indicate $P < 0.05$, 0.01, and 0.001 between KO mice treated with LPS and saline, respectively. $n = 7-9$.

2. LPS induced stress hormone release and anorexia unaffected in IL-1R1 KO mice

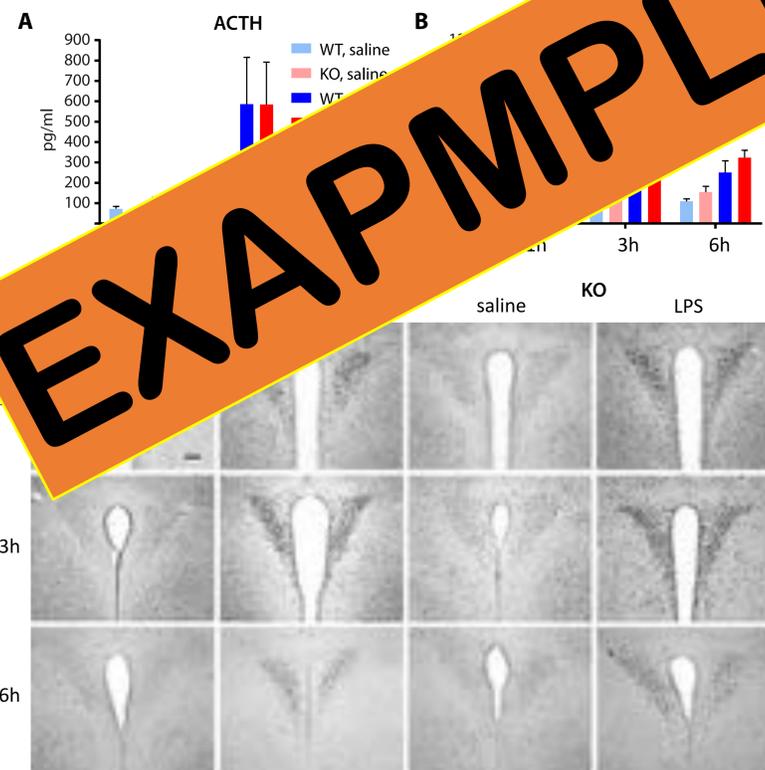


Figure 2. HPA-axis response to intraperitoneally injected LPS in WT and global IL-1R1 KO mice. A: ACTH in plasma. B: Corticosterone in plasma. C: Expression of Fos protein in the paraventricular nucleus of the hypothalamus. Scale bar = 100 μ m. In (A) and (B), $n = 5-6$ in the saline treated groups, and $n = 7-9$ in the LPS treated groups.

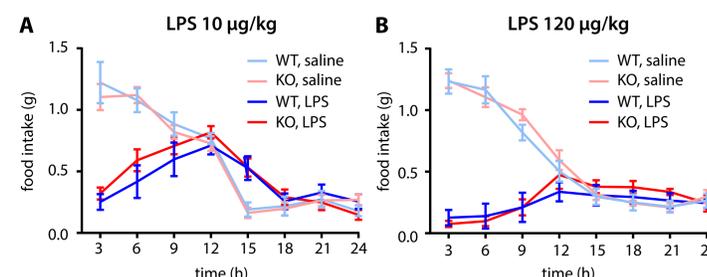


Figure 3. Food intake during 24 h in WT and global IL-1R1 KO mice injected intraperitoneally with a low (A) and a moderate dose (B) of LPS or saline. $n = 7-10$.

3. Neither TNF α inhibition nor IL-6 receptor blockade abolish the LPS induced febrile response in IL-1R1 KO mice

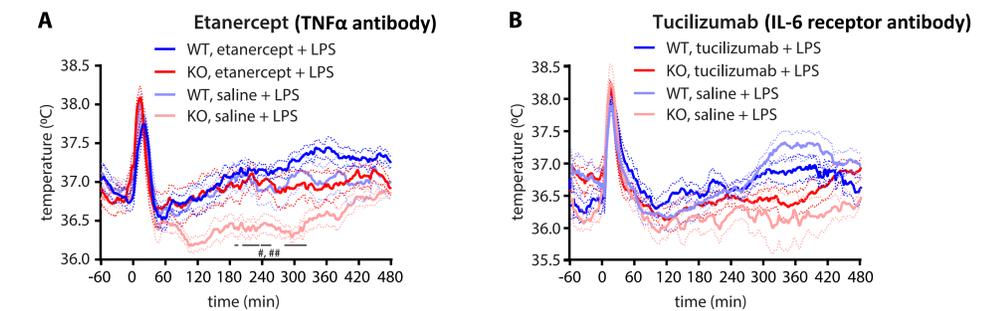


Figure 4. Effects of the TNF α antibody etanercept (A) and the IL-6 receptor antibody tucilizumab (B) on the febrile response to LPS. In (A), #, and ## indicate $P < 0.05$, and 0.01, between LPS injected global IL-1R1 KO mice treated with etanercept and saline, respectively. $n = 8-12$ in (A), and 6-8 in (B). Solid lines and dotted lines indicate the mean SEM.

4. Endothelial cell deletion of IL-1R1 attenuates LPS induced fever

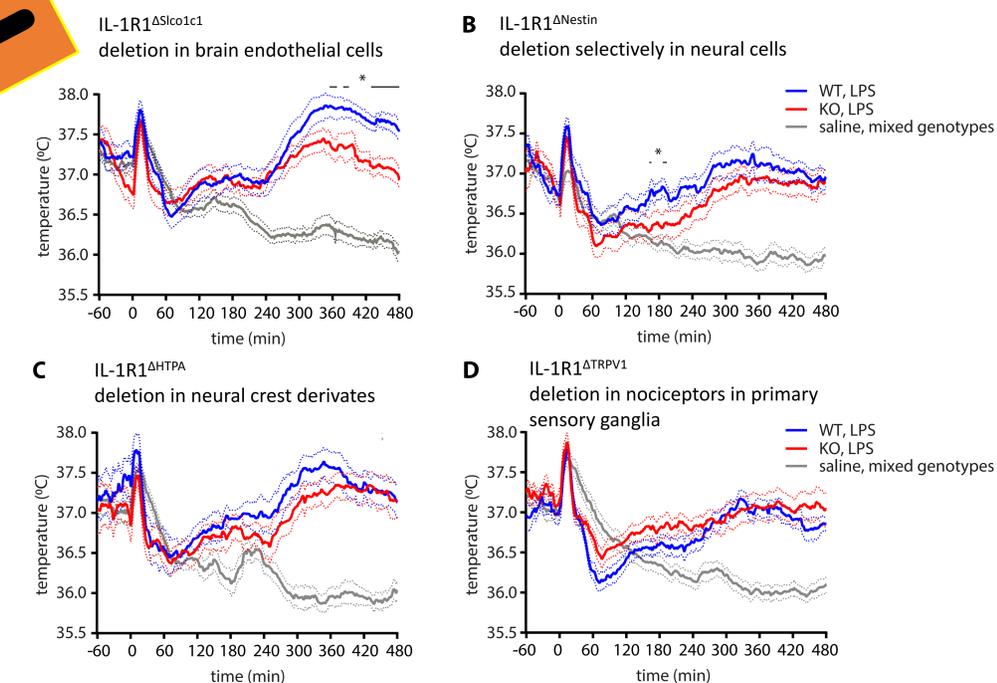


Figure 6. Febrile response to LPS in mice with IL-1R1 deletion selectively in (A) brain endothelial cells, (B) neural cells, (C) neural crest derivatives, and (D) nociceptive C-fibers. WT mice are IL-1R1^{fl/fl} littermates. * indicates $P < 0.05$ between WT mice and IL-1R1 KO mice. $n = 7-16$ for LPS treated animals; $n = 12-30$ for saline treated animals.

Summary

- IL-1R1 deletion reduces LPS-induced fever but not anorexia or HPA-axis activation
- Neither TNF α nor IL-6 receptors account for the febrile response in IL-1R1 KO mice
- IL-1R1s on brain endothelial cells are involved in the febrile response to LPS
- IL-1R1s on neural cells or peripheral nerves are not critical for LPS induced fever